

Sexually Antagonistic Variation and the Evolution of Dimorphic Sexual Systems

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ABSTRACT: Multicellular Eukaryotes use a broad spectrum of sexual reproduction strategies, ranging from simultaneous hermaphroditism to complete dioecy (separate sexes). The evolutionary pathway from hermaphroditism to dioecy involves the spread of sterility alleles that eliminate female or male reproductive functions, producing unisexual individuals. Classical theory predicts that evolutionary transitions to dioecy are feasible when female and male sex functions genetically trade off with one another (allocation to sex functions is sexually antagonistic) and rates of self-fertilization and inbreeding depression are high within the ancestral hermaphrodite population. We show that genetic linkage between sterility alleles and loci under sexually antagonistic selection significantly alters these classical predictions. We identify three specific consequences of linkage for the evolution of dimorphic sexual systems. First, linkage broadens conditions for the invasion of unisexual sterility alleles, facilitating transitions to sexual systems that are intermediate between hermaphroditism and dioecy (androdioecy and gynodioecy). Second, linkage elevates the equilibrium frequencies of unisexual individuals within androdioecious and gynodioecious populations, which promotes subsequent transitions to full dioecy. Third, linkage dampens the role of inbreeding during transitions to androdioecy and gynodioecy, making these transitions feasible in outbred populations. We discuss implications of these results for the evolution of dimorphic reproductive systems and sex chromosomes.

Keywords: dioecy, intralocus sexual conflict, linkage disequilibrium, sexual system, sexual dimorphism, sex chromosome evolution.

Introduction

Multicellular Eukaryotes use a diverse array of strategies for sexual reproduction (Bachtrog et al. 2014). At one end of

the spectrum, simultaneous hermaphrodites express both female and male reproductive structures (sex functions) and reproduce by outcrossing, self-fertilization, or a combination of both (mixed mating). At the other end of the spectrum, dioecious (or gonochoristic) species include separate females and males—usually at similar frequencies—that typically show pronounced sex differences in morphology, behavior, physiology, and life history (Andersson 1994). Flowering plants are particularly diverse in the range of sexual systems that they exhibit, which include hermaphroditism, dioecy, and nearly every possible state in between (Darwin 1877; Westergaard 1958; Bawa 1980; Sakai and Weller 1999; Barrett 2010; Bachtrog et al. 2014; Renner 2014; for animals, see Jarne and Auld 2006).

Dioecy has evolved repeatedly in plants from an ancestral state of functional hermaphroditism—an observation that has inspired much research into the evolutionary mechanisms that might drive such transitions (Westergaard 1958; Sakai and Weller 1999; Charlesworth 2006; Bachtrog et al. 2014; Renner 2014; Goldberg et al. 2017; Käfer et al. 2017). Theory and data agree that transitions from hermaphroditism to dioecy are likely to follow a two-step evolutionary pathway. In the first step, a nuclear unisexual sterility mutation spreads within a hermaphrodite population, resulting in the evolution of gynodioecy (a population composed of hermaphrodites and unisexual females) or androdioecy (hermaphrodites and males; Charlesworth and Charlesworth 1978*a*, 1978*b*). During the second step, the alternative sterility allele type—a female-sterility allele in a gynodioecious population or a male-sterility allele in an androdioecious population—spreads to complete the transition to full dioecy (Westergaard 1958; Charlesworth and Charlesworth 1978*a*, 1978*b*; Charlesworth 2006; Charlesworth and Willis 2009; Käfer et al. 2017). In these scenarios, unisexuality can arise through the invasion of mutations that individually cause the complete loss of an ancestral sex function in hermaphrodite individuals or through the spread of multiple alleles that cumulatively cause unisexuality (Charlesworth and Charlesworth 1978*a*, 1978*b*; Morgan 1992*a*, 1992*b*; Seger and Eckhart 1996;

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Charlesworth 1999). Current data on the evolution of plant sex determination systems are largely consistent with this two-step scenario (Westergaard 1958; Charlesworth 2002, 2006; Renner 2014; Ashman et al. 2015).

Classical theory has shown that transitions to dioecy are most likely to occur when two conditions are met (Lewis 1941; Lloyd 1975, 1976; Charlesworth and Charlesworth 1978a; Morgan 1992a; Seger and Eckhart 1996). First, female and male sex functions in hermaphrodites must genetically trade off against one another (e.g., through allocation trade-offs). Second, rates of self-fertilization and inbreeding depression in the ancestral hermaphrodite population must be high enough to promote the spread of unisexual sterility alleles that initiate the transition to dioecy. This theory predicts that gynodioecy, rather than androdioecy, represents the more likely intermediate state between hermaphroditism and dioecy (Lloyd 1975; Charlesworth and Charlesworth 1978a; Seger and Eckhart 1996; Käfer et al. 2017; for comprehensive reviews of relevant theory, see Charlesworth 1999, 2006). Finally, the second sterility mutation to invade the population is predicted to establish in tight linkage with the locus segregating for the first sterility allele (the linkage constraint of Charlesworth and Charlesworth 1978a). The establishment of complementary sterility mutations, under tight linkage and in repulsion phase, sets the stage for the subsequent evolution of heteromorphic sex chromosomes (e.g., a gene-rich X and degenerate Y; Bull 1983, pp. 265–269; Rice 1987; Qiu et al. 2013; reviewed in Charlesworth 2002; Bachtrog 2006).

One strength of the theory outlined above is that it makes clear and testable predictions about the types of species that are most likely to transition to dioecy. According to the theory, hermaphrodite lineages with high rates of self-fertilization are particularly likely to transition to dioecy through an intermediate gynodioecious state (Charlesworth and Charlesworth 1978a; Charlesworth 1999; Käfer et al. 2017). Yet, while some studies of single species support this long-standing prediction (reviewed in Webb 1999; Dufay and Billard 2012), empirical evidence for an association between dioecy and selfing across taxa remains equivocal (Charlesworth 1985, 2006; Renner 2014). The mismatch between the theoretical predictions and empirical patterns suggests that additional biological factors are likely to influence evolutionary transitions to dioecy, potentially overriding the facilitating effects of self-fertilization and inbreeding depression for the evolution of dioecy.

Here we extend population genetics theory for the evolution of dioecy by showing that genetic linkage of unisexual sterility alleles to a sexually antagonistic (SA) locus facilitates the initial step in the pathway to full dioecy (i.e., transitions from hermaphroditism to androdioecy or gynodioecy) and inverts the relation between self-fertilization and dioecy relative to predictions of classical theory. SA loci

refer to sites in the genome where fitness effects of genetic variation trade off between females and males or between hermaphrodite sex functions (see Gregorious 1982; Morgan 1992a, 1992b; Jordan and Connallon 2014; Olito 2017). Our models differ from the classical theories for the evolution of dioecy (Charlesworth and Charlesworth 1978a) by allowing unisexual sterility mutations to essentially hitchhike with genetically linked SA alleles, and vice versa. Linkage to an SA locus expands the range of conditions where unisexuals will spread, elevates the equilibrium frequencies of unisexuals in the population, and promotes evolutionary transitions to dioecy most readily in predominantly outcrossing populations. These findings have parallels in recent theory on the evolutionary consequences of multilocus SA genetic variation (Connallon and Clark 2010; Patten et al. 2010; Úbeda et al. 2010; Connallon and Jordan 2016; Olito 2017), and they suggest a new role for SA genetic variation in the evolutionary origins of new sex chromosome systems.

Models

Building on previous theory, we model evolutionary transitions from hermaphroditism to androdioecy and gynodioecy via the invasion of unisexual nuclear sterility alleles that cause complete male or complete female sterility, respectively (Charlesworth and Charlesworth 1978a). We separately consider the invasion of completely dominant and completely recessive sterility alleles during transitions to androdioecy and gynodioecy, leading to a total of four population genetic models. These scenarios mirror classical theory for the invasion of unisexuals in hermaphrodite populations (Charlesworth and Charlesworth 1978a), providing clear points of contrast. Below, we provide a full description of the simplest model: the evolution of gynodioecy via invasion of a dominant male-sterility mutation. We then briefly outline essential differences in the other three models, with reference to the first. Full details for all four models are provided in appendix A (apps. A–F are available online), and simulation code is available at <https://github.com/colin-olito/dioecySA>.¹

Gynodioecy

Consider a genetic system involving two diallelic autosomal loci, **A** (with alleles A , a) and **M** (with alleles M_1 , M_2), that recombine at rate r per meiosis. The **A** locus is under sexually antagonistic selection, with the A allele female beneficial and male deleterious and the a allele male beneficial and female deleterious. At the **M** locus, the M_1 allele has a relative fitness of one in both sexes, while the M_2 allele

1. Code that appears in *The American Naturalist* is provided as a convenience to readers. It has not necessarily been tested as part of peer review.

causes sterility through the male sex function and is completely dominant to the M_1 allele. Compared to hermaphrodites, female unisexuals reallocate resources that are normally invested in the male sex function (e.g., to pollen production) toward ovule production (Lloyd 1975, 1976; Charlesworth and Charlesworth 1978a), with k representing the proportional increase in ovule production by unisexual females relative to hermaphrodites. The population is initially hermaphroditic (fixed for M_1); evolution of gynodioecy requires the invasion of the M_2 allele into the population. Cytoplasmic male sterility may also play a role in the evolution of gynodioecy, but transitions to full dioecy require the subsequent invasion of nuclear sterility alleles (e.g., Lewis 1941; Frank 1989; Charlesworth 2002, 2006).

Among offspring of hermaphrodites, a fixed proportion C are produced by self-fertilization, and the remainder $(1 - C)$ are produced by outcrossing. Unisexual females cannot self-fertilize. Individuals produced by self-fertilization suffer from inbreeding depression, with δ representing the decreased probability of survival to reproductive maturity of self-fertilized zygotes relative to those produced by outcrossing. Generations are discrete, and the life cycle proceeds as follows: birth \rightarrow differential survival due to inbreeding depression \rightarrow selection on reproductive success through male and female sex functions \rightarrow random fertilization.

Let x_i and y_i denote the frequencies of the four possible haplotypes [AM_1, AM_2, aM_1, aM_2] in female and male gametes, respectively. An individual carrying haplotypes i and j has fitness of w_{ij}^f through the female sex function and w_{ij}^m through the male sex function; fitness through each sex function is multiplicative between the **A** and **M** loci (table 1). We assume that there are no parent-of-origin effects on fitness.

To track evolutionary changes in genotype frequencies, we partitioned genotypes by whether they were produced by self-fertilization or by outcrossing. The approach yields a system of 20 general recursion equations (10 genotypes \times two modes of reproduction; see app. A). Although more compact approaches have been used for generating recursions in previous models (e.g., Charlesworth and Charlesworth 1978a, 2010), we used the following expanded expressions to clarify the transmission pathway and assumptions underlying each of our models. As shown below, our models reproduce the classical results in special cases where there is only selection at loci segregating for sterility alleles.

Let F_{ij} represent the zygotic frequency of outcrossed genotypes carrying haplotypes i and j and G_{ij} represent the frequency of the same genotype among zygotes produced by self-fertilization. When M_2 is completely dominant to M_1 , the recursions for outcrossed zygotes in the next generation simplify to

$$\begin{aligned} F'_{11} &= (1 - S)(x_1y_1), & F'_{23} &= (1 - S)(x_2y_3), \\ F'_{12} &= (1 - S)(x_2y_1), & F'_{24} &= 0, \\ F'_{13} &= (1 - S)(x_1y_3 + x_3y_1), & F'_{33} &= (1 - S)(x_3y_3), \\ F'_{14} &= (1 - S)(x_4y_1), & F'_{34} &= (1 - S)(x_4y_3), \\ F'_{22} &= 0, & F'_{44} &= 0, \end{aligned} \tag{1}$$

where S is the proportion of all ovules in the population that are self-fertilized (S is proportional to the selfing rate, C , and takes into account selection on ovule production and the fact that not all genotypes produce pollen; see app. A for details). Note that $y_2 = y_4 = 0$ because these male gametic haplotypes cannot be produced when M_2 is a dominant male-sterility allele. Among zygotes formed by self-fertilization, the genotypic frequencies in the next generation are

Table 1: Fitness expressions for diploid adults prior to reproduction for the model of a dominant male-sterility mutation, where w_{ij}^f denotes fitness effects through the female sex function and w_{ij}^m through the male sex function

Haplotype	AM_1	AM_2	aM_1	aM_2
Female sex function:				
AM_1	1	$(1 + k)$	$(1 - h_f s_f)$	$(1 - h_f s_f)(1 + k)$
AM_2	...	$(1 + k)$	$(1 - h_f s_f)(1 + k)$	$(1 - h_f s_f)(1 + k)$
aM_1	$(1 - s_f)$	$(1 - s_f)(1 + k)$
aM_2	$(1 - s_f)(1 + k)$
Male sex function:				
AM_1	$(1 - s_m)$	0	$(1 - h_m s_m)$	0
AM_2	...	0	0	0
aM_1	1	0
aM_2	0

Note: Rows and columns indicate the i th and j th gametic haplotypes, respectively. The lower triangle of each matrix is the reflection of the upper triangle and is omitted for simplicity and consistency with the $i \geq j$ row/column indexing used throughout the article.

$$\begin{aligned} G'_{11} &= S\left(o_{11}^S + \frac{o_{13}^S}{4}\right), \\ G'_{13} &= S\left(\frac{o_{13}^S}{2}\right), \\ G'_{33} &= S\left(o_{33}^S + \frac{o_{13}^S}{4}\right), \end{aligned} \quad (2)$$

where o_{ij}^S are functions describing the proportional contribution of each genotype to self-fertilized ovules (see app. A). All $G'_{ij} = 0$, where $ij \neq [11, 13, 33]$.

The basic form of the recursions does not change when M_2 is recessive, but there are two notable differences. First, because only M_2M_2 individuals are unisexual females, none of the recursions reduce to zero. Second, in contrast to the gynodioecy model with dominant M_2 allele, the recombination rate parameter (r) enters into the G'_{ij} recursions and the functions x_i and y_i in the recessive M_2 model (see app. A).

Androdioecy

Androdioecy evolves when an M_2 allele causing female sterility invades a hermaphrodite population. In the androdioecy model, the reproductive compensation term, k , now describes the increase in pollen production by males relative to hermaphrodites. When the M_2 allele is dominant (i.e., M_2 carriers are male), the fitness expressions w_{ij}^f and w_{ij}^m resemble those described in table 1, except that the **M** locus affects the female rather than the male sex function.

Recursions for the androdioecy models are similar to those for gynodioecy, with a few key differences. When M_2 is dominant, the recombination rate drops out of the expressions for haplotype frequencies in ovules (x_i ; $x_2 = x_4 = 0$, as in the gynodioecy model with dominant M_2 allele), whereas haplotype frequencies in pollen are partly dependent on recombination (functions for y_i include r terms). The genotypic frequencies in the next generation among zygotes formed by outcross fertilization, F'_{ij} , are otherwise identical to equation (1), except that heterozygotes at the **M** locus do not produce ovules. The form of the G'_{ij} recursions remains unchanged from equation (2) (see app. A).

When the M_2 (female-sterility) allele is recessive, only M_2M_2 homozygotes are unisexual males. The form of the recursions is similar to the case of gynodioecy with a recessive M_2 allele but with the **M** locus now affecting female sterility. Expressions for the ovule and pollen haplotype frequencies, x_i and y_i , partially depend on the recombination rate (see app. A).

Analyses

Because they are analytically tractable for linear stability analyses, we used the dominant unisexual sterility allele

models to determine conditions that favor the spread of sterility and SA alleles (see the introduction).

Following previous SA theory (e.g., Kidwell et al. 1977; Prout 2000; Jordan and Connallon 2014), we focused our analyses on the simplest case of codominant (i.e., additive) expression of SA alleles at locus **A** ($h_m = h_f = 1/2$ in table 1). Additional results exploring the effects of dominance reversals at the **A** locus ($h_m, h_f < 1/2$) are presented in appendix C. These additional results are similar to those of the additive scenario, except that the invasion conditions for both SA and sterility alleles are more permissive (Fry 2009; Jordan and Connallon 2014; Olito 2017).

We first examined the case where both loci are initially monomorphic (M_1 is fixed at the **M** locus; A or a is fixed at the **A** locus). To identify conditions for the invasion of derived alleles at each locus (individually or in combination), we carried out a linear stability analysis of each monomorphic condition corresponding to $F_{11} = G_{11} = 1$ when A is initially fixed and to $F_{33} = G_{33} = 1$ when a is initially fixed. An equilibrium is unstable and a derived haplotype will invade the population when the leading eigenvalue of the Jacobian matrix of the system of recursions is greater than one, $\lambda_L > 1$; Otto and Day 2007). For the models involving dominant unisexual sterility alleles, each analysis yields three analytically tractable candidate-leading eigenvalues. The first two candidate eigenvalues (λ_A and λ_M) describe conditions for invasion of the derived allele at each locus singly. The third candidate eigenvalue describes the condition for invasion of a haplotype carrying both derived alleles (λ_{AM}). Mathematica code for our analytic results are presented in appendix F.

We next explored how SA polymorphism influences the invasion of unisexual sterility alleles into a population of hermaphrodites. For this scenario, we evaluated whether the M_2 allele can invade a population at deterministic polymorphic equilibrium at the **A** locus, with M_1 initially fixed at the **M** locus. As we note in the discussion, the assumption of polymorphic equilibrium at the SA locus is made for convenience in the stability analyses and is not a requirement for invasion of the unisexual sterility alleles. For the initial conditions of SA polymorphisms, we used the exact single-locus equilibria from Kidwell et al. (1977) for outcrossing populations with additive SA effects. We used approximate single-locus equilibria (see Jordan and Connallon 2014; Connallon and Jordan 2016) for cases of partial selfing and/or nonadditive effects of SA alleles (the expressions for single-locus SA polymorphism are provided in app. B). The approximations compare well against exact numerical results when selection at the SA locus is modest to weak ($s_p, s_m \leq 0.5$; Jordan and Connallon 2014; Connallon and Jordan 2016; Olito 2017).

Linear stability analyses are inconclusive when sterility alleles are recessive. Yet the evolutionary dynamics of recessive

sive sterility alleles are of interest because such alleles are probably more common than dominant ones (Charlesworth and Charlesworth 1978a). We therefore carried out deterministic simulations of the exact recursions to find the equilibrium frequencies for each model of recessive unisexual sterility. To assess the impact of linkage on the polymorphic frequencies of unisexuals and hermaphrodites, we compared equilibrium frequencies from our models with the corresponding single-locus equilibria from Charlesworth and Charlesworth (1978a; denoted \hat{Z}). For these comparisons, we focus on relatively tight linkage between **A** and **M** ($r \leq 0.1$). Predictions from one- and two-locus models converge under loose linkage (high r).

In single-locus models of gynodioecy and androdioecy, the equilibrium frequency of unisexuals is determined by the reproductive compensation term k and the compound parameter $C\delta$ (Charlesworth and Charlesworth 1978a), where C and δ refer to the rate of selfing and the severity of inbreeding depression, respectively. If inbreeding depression is caused primarily by recessive deleterious mutations, as current data suggest (Charlesworth and Willis 2009), C and δ should negatively covary, as increased selfing purges deleterious recessives (we note, however, that other processes also contribute to observed patterns of inbreeding depression, e.g., Crnokrak and Barrett 2002; Charlesworth et al. 2007; Charlesworth and Willis 2009; Hedrick and Garcia-Dorado 2016). For simplicity, we incorporated negative covariance into our simulations by constraining inbreeding depression to follow a linear declining function of the selfing rate: $\delta = \delta^*(1 - C/2)$, where δ^* represents the hypothetical severity of inbreeding depression for an outcrossing population. We set $\delta^* = 0.8$ for all simulations, which resulted in levels of inbreeding depression that are consistent with empirical data (e.g., fig. 2 in Husband and Schemske 1996). We emphasize that our intention is not to formally model the evolution of inbreeding depression but to capture effects of purging deleterious recessives in partially selfing populations and thereby explore biologically meaningful parameter space for the evolution of androdioecy and gynodioecy. In fact, a variety of detailed genetic models give rise to a nonlinear relation between inbreeding depression and selfing (e.g., Ohta and Cockerham 1974; Charlesworth 1985; Lande and Schemske 1985; Roze 2015; Garcia-Dorado 2017; Lande and Porcher 2017). As we show in appendix E, such nonlinear expressions for δ as a function of C yield similar results to the linear expression described above.

To take into account the fact that C and δ also influence the maintenance of SA polymorphism (Jordan and Connallon 2014; Olito 2017), we ran simulations using values of s_f and s_m that correspond to equal equilibrium frequencies for the two SA alleles ($p = q = 1/2$). Thus, our simulations explore whether unisexual sterility alleles will invade

hermaphroditic populations at polymorphic equilibrium with equal frequencies of male- and female-beneficial SA alleles. We reiterate, however, that such balanced SA polymorphisms are not strictly necessary for sterility alleles to invade (see “Discussion”).

Results

Invasion of Unisexuals into Monomorphic Populations

Gynodioecy. We begin with the simplest scenario for the evolution of gynodioecy: invasion of a dominant male-sterility allele, M_2 , into a hermaphrodite population that is initially monomorphic for the SA locus (fixed for the AAM_1M_1 or the aaM_1M_1 genotype, which includes cases where selection favors the fixation of one of the SA alleles).

A new haplotype can spread within the population under three conditions. First, when the single-locus invasion criteria for the rare SA allele is met ($\lambda_A > 1$; Kidwell et al. 1977; Jordan and Connallon 2014; Olito 2017), selection will favor its invasion. This single-locus invasion condition applies broadly to cases of additive and nonadditive effects of SA alleles.

Second, a male-sterile allele can invade in the absence of linkage to an SA allele when the classic single-locus criterion for the evolution of gynodioecy is met ($\lambda_M > 1$):

$$k > 1 - 2C\delta \quad (3)$$

(see eq. [4] of Charlesworth and Charlesworth 1978a). Recall that k is the increased ovule production in females relative to hermaphrodites, and δ represents the strength of inbreeding depression in self-fertilized relative to outcrossed individuals. For convenience, we define \hat{k} as the threshold level of reproductive compensation for invasion of a unisexual in the single-locus model; \hat{k} is equal to the right-hand side of equation (3), and $k > \hat{k}$ is the single-locus condition for invasion.

Third, a mutant haplotype bearing both the male-sterile allele and the derived allele at the SA locus can invade when $\lambda_{AM} > 1$. This condition for invasion takes the same basic form as equation (3), in which the minimum value of k that is required for invasion is a decreasing function of $C\delta$ (see app. F). When the population is initially fixed for the female-beneficial SA allele (A), the haplotype invasion condition is more restrictive (requiring larger k) than the single-locus invasion condition (eq. [3]). In contrast, when the population is initially fixed for the male-beneficial allele (a), the condition for invasion of the derived haplotype is more permissive, provided the recombination rate is low relative to the strength of selection at the SA locus:

$$r < \frac{s_f}{2 - s_f} \quad (4)$$

Equation (4) shows that tight linkage, strong SA selection, or both, facilitates the invasion of unisexual females and the evolution of gynodioecy. Nevertheless, strong selection is not required for linkage to have a pronounced effect on the conditions for invasion of unisexuals. For example, when $s_j \leq 0.1$, the threshold k for unisexual invasion decreases by up to ~15% relative to single-locus predictions (fig. 1A; see also app. D, fig. D1; figs. D1–D16, E1 are available online). Stronger SA selection decreases the threshold for invasion of a unisexual sterility allele. Importantly, the above result holds whether the linked female-beneficial SA allele is ultimately fixed or is maintained as a balanced polymorphism.

Linkage to a male-sterility allele also expands conditions for the invasion of female-beneficial SA alleles. Under obligate outcrossing, conditions for invasion of a derived haplotype with the female-benefit SA allele and male-sterility allele (AM_2 ; corresponding to $\lambda_{AM} > 1$) are more permissive than the single-locus SA invasion condition when equation (4) is satisfied. Under partial selfing, the invasion condition for

the same derived haplotype becomes more permissive than the single-locus prediction when

$$r < \frac{s_m(1-C)}{2+s_m+C(2-s_m-4\delta)}. \quad (5)$$

Equation (5) shows that selfing dampens the linkage-induced expansion of parameter space for invasion of female-beneficial SA alleles. This dampening effect occurs because increased selfing biases evolution in favor of the female-beneficial alleles (Olito 2017); the bias leaves less parameter space available for linkage to further expand the conditions where female-beneficial SA alleles can invade. Nevertheless, tight linkage can still aid invasion of female-beneficial alleles provided there is some degree of outcrossing among hermaphrodites.

Androdioecy. We identified conditions for the evolution of androdioecy by analyzing the stability of a population initially fixed for the AAM_1M_1 or the aaM_1M_1 genotype. We now follow the evolution of a dominant female-sterility al-

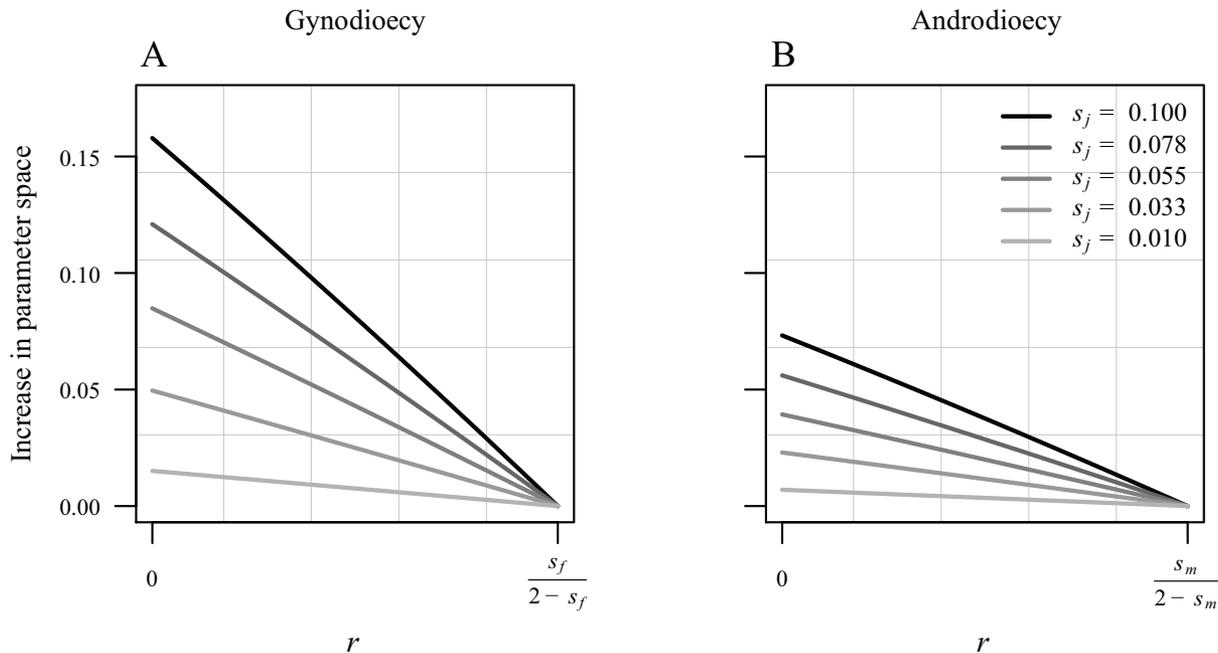


Figure 1: Effect of linkage between the A and M loci on the invasion of unisexual females (A) and males (B) into hermaphrodite populations. Plots show the increase in parameter space (calculated as a proportion of the parameter space where eq. [3] is satisfied) for the invasion of double mutants (i.e., based on λ_{AM}) compared with the single-locus invasion criteria (i.e., based on λ_M ; see eqs. [3] and [6] for the models of gynodioecy and androdioecy, respectively) for different selection intensities as a function of the recombination rate between the two loci. We vary the recombination rate between zero and the threshold where the one- and two-locus invasion conditions become equivalent (eq. [4]). For the model of gynodioecy, we first solved $\lambda_{AM} > 1$ for k (denoted \hat{k}_{AM} for convenience) and then integrated both \hat{k}_{AM} and \hat{k} (eq. [3], right-hand side) over $C = [0, 1]$ and $\delta = [0, 1]$ for the specified values of r and s_j (where $j \in \{f, m\}$). The increase in parameter space (decrease in \hat{k}_{AM} relative to \hat{k}) was then calculated by subtracting the resulting volume for \hat{k}_{AM} from that of \hat{k} and dividing by the latter. Thus, the parameter space where unisexual females can invade is increased in the two-locus model because linkage reduces \hat{k}_{AM} relative to \hat{k} across biologically plausible values of C and δ (see fig. D1). Because the integral of equation (6) is undefined when $C = 1$, we restricted the integration to include high, but not complete, selfing rates $C = [0, 0.9]$ for the model of androdioecy; comparisons between the models should therefore be made with caution.

lele (M_2) and SA alleles with additive effects on the sex functions. Analysis of the first eigenvalue (λ_A) recovers the general, single-locus invasion criterion for a derived SA allele (see above). Analysis of the second eigenvalue (λ_M) yields the familiar single-locus condition for invasion of unisexual males into a hermaphrodite population:

$$k > \frac{1 + C(1 - 2\delta)}{(1 - C)} \quad (6)$$

(see eq. [8] of Charlesworth and Charlesworth 1978a). As before, we define \hat{k} as the threshold level of reproductive compensation for unisexual invasion in the single-locus model, with \hat{k} equal to the right-hand side of equation (3) (for the gynodioecy model) or equation (6) (for the androdioecy model), as appropriate.

Haplotypes bearing derived alleles at both loci can invade when $\lambda_{AM} > 1$, where invasion conditions are once again dependent on the SA genotype that is initially fixed in the ancestral hermaphrodite population (see app. F). Mirroring the results of the gynodioecy model, the conditions for invasion are more restrictive than the single-locus condition (eq. [6]) when the population is initially fixed for the male-beneficial allele (a). Conditions for invasion are more permissive when the female-beneficial allele (A) is initially fixed. Replacing s_f with s_m in equation (4) yields the condition under which linkage to the SA locus promotes the invasion of unisexual males (i.e., relative to eq. [6]). The expansion of parameter space for invasion of the two-locus derived haplotype is smaller in the androdioecy model compared to the gynodioecy model (fig. 1B). Again, this condition holds whether or not the invading male-beneficial allele will ultimately fix.

Linkage to a female-sterility allele also facilitates the invasion of male-beneficial SA alleles. With s_m substituted for s_f , equation (4) provides the condition where linkage to a female-sterility allele expands the invasion condition of a male-beneficial SA allele in an obligate outcrossing population. Under arbitrary selfing in hermaphrodites, linkage to a female-sterility locus expands the invasion conditions of male-beneficial SA alleles when

$$r < \frac{s_f(1 + C(1 - 2\delta))}{2 - 2C + s_f(1 + C(1 - 2\delta))}. \quad (7)$$

In contrast to the gynodioecy model, where selfing dampens the expansion of parameter space for the linked invasion of female-beneficial SA alleles (eq. [5]), self-fertilization expands conditions for invasion of male-beneficial SA alleles that are linked to female-sterility alleles in the androdioecy model. For example, the right-hand side of equation (7) increases with both C and s_f such that, for $C > 1/3$, there is always some expansion of the parameter space where male-beneficial alleles can invade relative to the single-locus ex-

pectation, even under free recombination ($r = 1/2$; see also Olito 2017).

Invasion of Unisexuals into Polymorphic Populations

Reproductive compensation, recombination, and selfing rates have similar effects on the invasion of sterility mutations into populations maintaining polymorphic SA alleles compared to populations where the SA locus is initially monomorphic (above). Linkage to a polymorphic SA locus expands the parameter conditions for the invasion of unisexuals relative to the classical single-locus conditions (fig. 2). In cases where reproductive compensation falls below the single-locus invasion threshold, unisexuals can still invade when the sterility mutation and SA locus are sufficiently linked. For example, under obligate outcrossing and tight linkage ($r = 0$), unisexuals can invade across ~69% of the parameter conditions where SA polymorphism is maintained (within the range $s_f, s_m \leq 0.5$), even if they suffer a 10% reduction in gamete production relative to the single-locus invasion criterion for unisexual sterility alleles (the right-hand side of eq. [3]; see fig. 2A, 2D). With smaller reductions in gamete production relative to the single-locus invasion criterion, unisexuals can invade across a greater fraction of polymorphic SA parameter space, even when linkage is relatively loose (e.g., when $r = 0.2$ and $k = 0.95 \times \hat{k}$, unisexuals will invade across ~38% of parameter conditions maintaining SA polymorphism).

With a small amount of selfing and high inbreeding depression in hermaphrodites ($C = 1/4, \delta = 0.8$), the breadth of parameter space where unisexuals can invade remains similar to the obligate outcrossing scenario (fig. 2B, 2E). However, under high selfing rates and low inbreeding depression ($C = 3/4, \delta = 0.2$), tighter linkage is required for invasion of male-sterility alleles during transitions to gynodioecy than for female-sterility alleles during transitions to androdioecy, when reproductive compensation falls below the single-locus invasion threshold (fig. 2C, 2F). That is, linkage to an SA locus has a stronger facilitating effect on transitions to androdioecy than for transitions to gynodioecy. As before, this contrasting effect of selfing between the gynodioecy and androdioecy models arises because selfing causes asymmetry in the evolutionary dynamics of SA alleles, which favors female-benefit relative to male-benefit alleles (Charlesworth and Charlesworth 1978a; Jordan and Connallon 2014; see app. D, figs. D2–D7, for examples). The evolutionary bias toward female-beneficial alleles creates greater scope for linkage to a male-beneficial SA allele to aid invasion of a female-sterility allele (resulting in androdioecy) than for linkage to a female-beneficial allele to aid the spread of a male-sterility allele (resulting in gynodioecy).

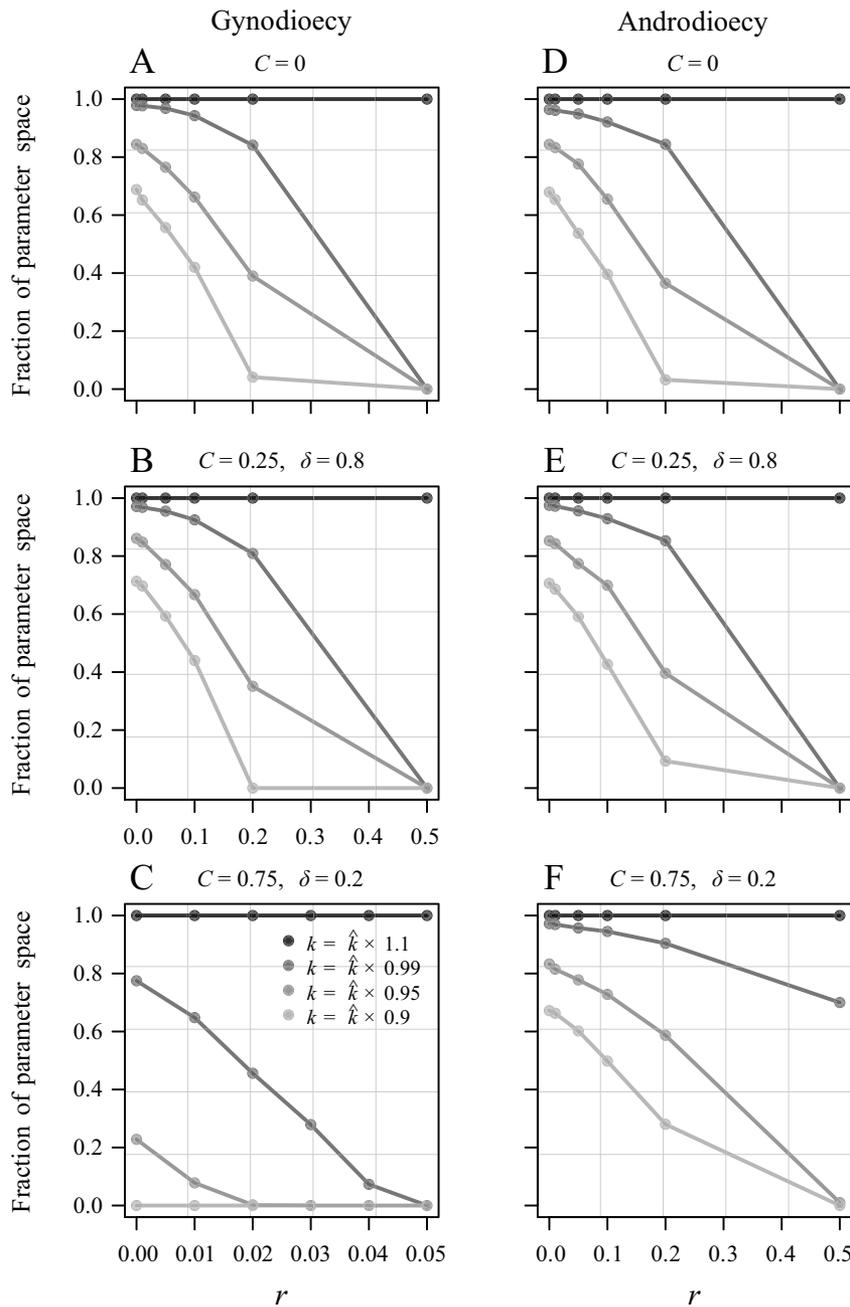


Figure 2: Invasion of unisexuals into populations with preexisting SA polymorphism. Plots show the fraction of parameter conditions maintaining single-locus SA polymorphism (within the range $0 < s_f, s_m \leq 0.5$) where a dominant sex-specific sterility allele at M can invade, assuming additive SA fitness effects at the SA A locus ($h_f = h_m = 1/2$), plotted as a function of the recombination rate r . Panels A–C show results from the model of gynodioecy via invasion of a male-sterility allele, while panels D–F show results for the model of androdioecy via invasion of a female-sterility allele. For each panel, results are shown for different values of reproductive compensation, k , chosen as a fraction of the single-locus invasion threshold for the unisexual sterility allele (\hat{k} , which is equal to the right-hand side of eq. [3] or eq. [6] for the models of gynodioecy and androdioecy, respectively). Note the different scale for the X-axis in C. To calculate the fraction of parameter space where unisexuals can invade populations initially polymorphic at the SA locus, we first drew 1,000 points uniformly distributed throughout polymorphic $s_f \times s_m$ parameter space (i.e., where the single-locus equilibrium frequency of the female-beneficial SA allele, \hat{q} , satisfied $0 < \hat{q} < 1$). The fraction of parameter space was calculated as the fraction of these 1,000 points at which either of the candidate-leading eigenvalues associated with invasion of unisexuals (λ_M, λ_{AM}) evaluated in excess of one.

SA genetic variation will facilitate the spread of loosely linked unisexual sterility alleles when selection at the SA locus is strong (larger values of s_m and s_f), and selection is biased toward the sex of the invading unisexual type. In other words, with incomplete linkage, the invasion of male-sterility alleles requires female-biased selection ($s_f < s_m$), while the invasion of female sterility requires male-biased selection ($s_f > s_m$; see app. D, figs. D2–D7).

*Equilibrium Frequencies of Unisexu-
als When Sterility Alleles Are Recessive*

Relative to classical single-locus predictions, linkage with an SA locus elevates the equilibrium frequencies of recessive sterility alleles and unisexu-als, consistent with results of the dominant sterility allele models. Indeed, when reproductive compensation matches or exceeds the threshold for unisexual invasion in the single-locus model (i.e., $k \geq \hat{k}$, where \hat{k} is equal to the right-hand side of eq. [3] or eq. [6] as appropriate), linkage to an SA locus can greatly increase the equilibrium frequencies of unisexu-als relative to single-locus predictions (Charlesworth and Charlesworth 1978a; figs. 3, D15, grayscale lines). The effect is strongest in pre-dominantly outcrossing populations and weakens with in-

creasing rates of selfing in hermaphrodites (recall that in-breeding depression also declines with selfing in our models; see “Models”). As the recombination rate between loci in-creases, unisexual equilibrium frequencies converge to pre-dictions of the single-locus models.

When reproductive compensation falls below the single-locus threshold for invasion ($k < \hat{k}$), the combination of tight linkage and outcrossing can still elevate the equi-librium frequencies of unisexu-als (fig. 4A, 4C, with the effect declining with the recombination rate, the rate of outcrossing, and the magnitude of reproductive compensation: fig. 4B, 4D). In these cases, the frequencies of males in the andro-dioecy model are more sensitive to reductions in k than for the equilibrium frequencies of females in the gynodioecy model. With lower reproductive compensation, equilibrium unisexual frequencies are highest for populations with inter-mediate selfing rates, reflecting a balance between two oppo-sing factors that influence the evolution of recessive sterility alleles: whereas selfing increases both the expression of uni-sexual phenotypes (by increasing the frequency of M_2M_2 homozygotes) and the opportunity for selection for rare uni-sexual sterility alleles, it also places a limit on the reproduc-tive success of unisexu-als, which can only reproduce by out-crossing.

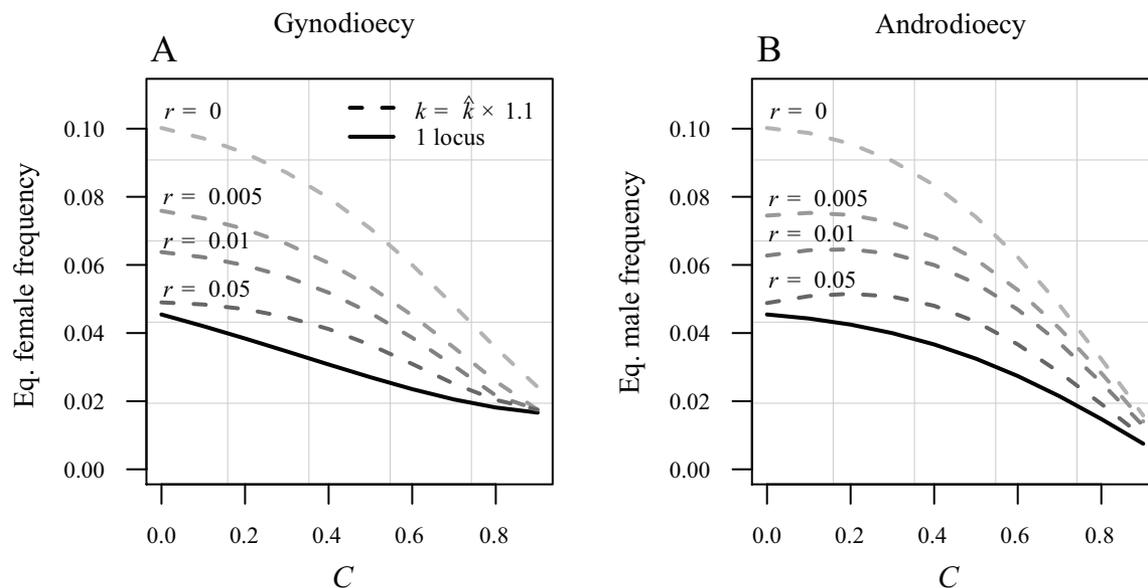


Figure 3: Effect of linkage on equilibrium frequencies of unisexual females (A) and males (B) when reproductive compensation is above the single-locus threshold for invasion of unisexual sterility allele (i.e., $k > \hat{k}$, where \hat{k} is equal to the right-hand side of eq. [3] or eq. [6], as appropriate). Results are shown for the models of androdioecy and gynodioecy via invasion of recessive unisexual sterility alleles, additive fitness effects at A ($h_f = h_m = 0.5$, using selection coefficients of $s_m = 0.1$ for the model of gynodioecy and $s_f = 0.1$ for the model of androdioecy), and inbreeding depression that follows $\delta = \delta^*(1 - C/2)$ (see “Models”). Plots illustrate the increase in equilibrium frequencies of unisexu-als predicted by our two-locus models (dashed grayscale lines) relative to the corresponding exact single-locus equilibrium frequencies (solid black line; \hat{Z} predicted by Charlesworth and Charlesworth 1978a). Results are shown for four different levels of recombination, highlighting that with weaker linkage, the two-locus predictions converge on those of the single-locus model.

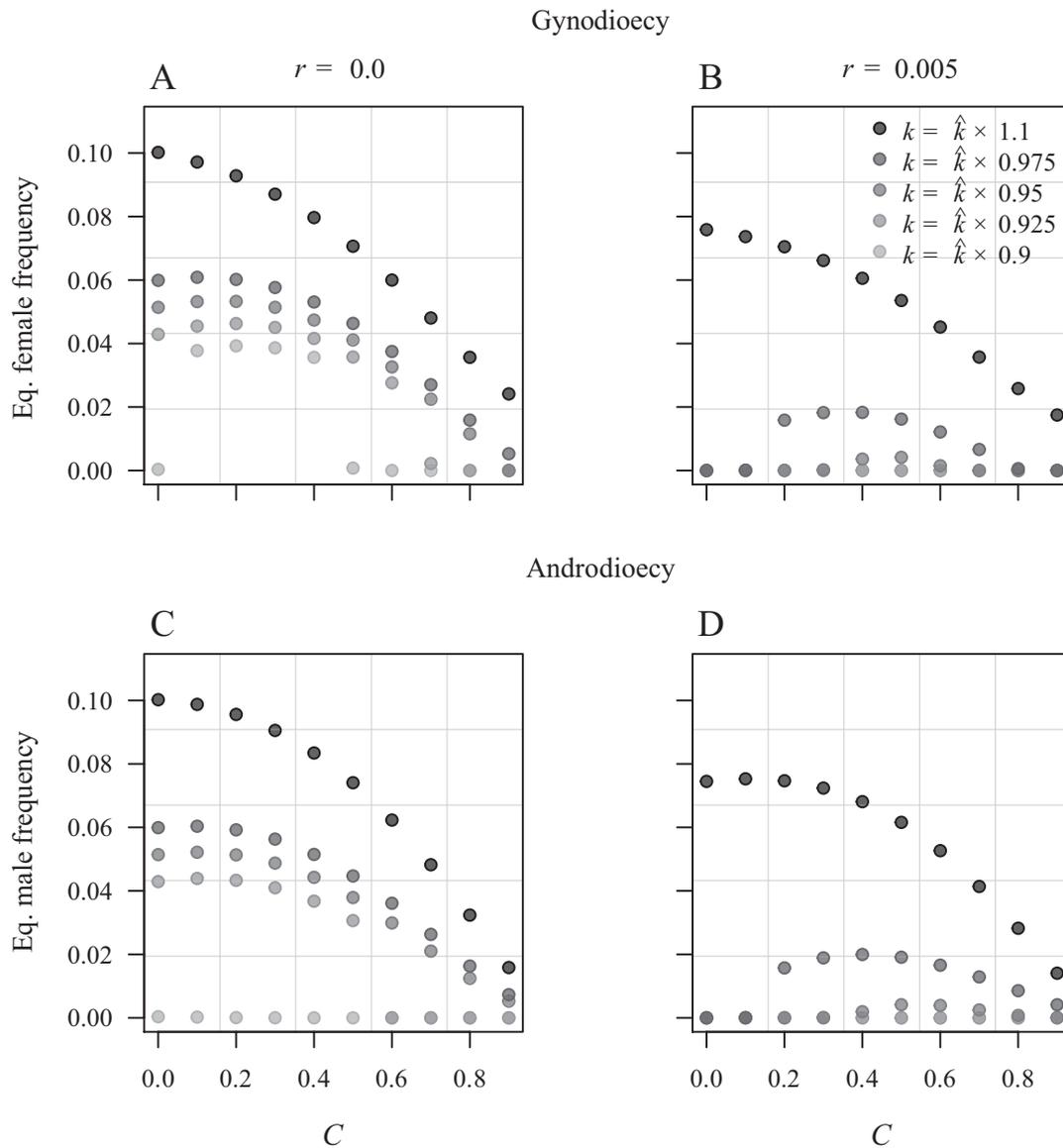


Figure 4: Equilibrium frequencies of unisexual females (A, B) and males (C, D) across a gradient of reproductive compensation (with values chosen as a fraction of the single-locus invasion threshold for M_2 (\hat{k} , which is equal to the right-hand side of eq. [3] or eq. [6] for the models of gynodioecy and androdioecy, respectively)). Results are shown for the models of androdioecy and gynodioecy via invasion of recessive unisexual sterility alleles, additive fitness effects at A ($h_f = h_m = 0.5$, using selection coefficients of $s_m = 0.1$ for the model of gynodioecy and $s_f = 0.1$ for the model of androdioecy), and inbreeding depression that decreases linearly with the selfing rate: $\delta = \delta^*(1 - C/2)$ (see “Models”). Plots show the equilibrium frequencies of unisexuals predicted by our models for five different levels of reproductive compensation, calculated as a fraction of the single-locus invasion criterion for M_2 defined by equations (3) and (6). Note that the single-locus equilibrium frequency of unisexuals always equals zero when $k < \hat{k}$. Hence, the lines corresponding to $k < \hat{k}$ illustrate how linkage among SA loci expands the parameter space where unisexual sterility alleles can invade beyond the predictions of the single-locus models.

Discussion

Our theoretical predictions have three implications for the evolution of mating systems and the genetic basis of separate sexes. First, compared to classical single-locus models of androdioecy and gynodioecy, linkage to an SA locus ex-

pands the range of conditions where unisexual sterility alleles are favored and elevates the equilibrium frequencies of invading unisexual individuals within androdioecious and gynodioecious populations. Second, the facilitating effect of linkage on the invasion of unisexuals is most pronounced in outcrossing hermaphrodite populations, suggesting that

correlations between the ancestral selfing rate and the evolution of separate sexes may be weaker than predicted by earlier theory, in which selfing promotes invasion of unisexuals (see Charlesworth and Charlesworth 1978*a*). Third, successfully invading unisexual sterility alleles, which initiate transitions to dioecy, are expected to preferentially accumulate in genomic regions harboring SA polymorphism—a prediction at odds with the prevailing view that sexual antagonism is more likely to follow rather than precede the evolution of separate sexes and of genetic sex determination (reviewed in Charlesworth et al. 2005; Bachtrog 2006). Below, we elaborate on the predictions of our models and suggest ways to test them.

SA Polymorphism and the Evolution of Dimorphic Sexual Systems

Classical theory predicts that conditions for the evolution of dioecy are likely to be restrictive (Lloyd 1975, 1976; Charlesworth and Charlesworth 1978*a*; Käfer et al. 2017). Unisexuals can invade a population when they compensate for the loss of one ancestral sex function by increasing gamete production through the other (e.g., unisexual females can compensate for the loss of the male sex function by producing more ovules than hermaphrodites do). We find that when sterility mutations arise on haplotypes that carry complementary SA alleles, the conditions for the spread of unisexual sterility require less compensation than predicted by previous theory (e.g., a male-sterility allele is more likely to spread when it arises on a haplotype carrying a female-beneficial SA allele). Intuitively, linkage to a complementary SA allele helps to offset the loss of a sex function caused by the sterility allele, thereby reducing the amount of reproductive compensation required for unisexual invasion relative to single-locus predictions (Charlesworth and Charlesworth 1978*a*). These results parallel recent multilocus models of sex-specific selection, which show that predominantly female-harming mutations tend to accumulate on haplotypes carrying male-benefit SA alleles and that male-harming mutations preferentially accumulate in linkage with female-beneficial SA alleles (Blackburn et al. 2010; Patten et al. 2010; Úbeda et al. 2010; Connallon and Jordan 2016). A corollary of this result is that conditions for the maintenance of SA polymorphism are also expanded by linkage to sex-specific sterility mutations. Hence, the invasion of new unisexual sterility mutations (i.e., during the initial step in the evolution of dioecy) should also promote the maintenance of polymorphism at nearby SA loci.

Balanced SA polymorphisms are not a prerequisite for linkage between an SA locus and a unisexual sterility mutation to facilitate transitions to androdioecy or gynodioecy. However, the amount of standing SA genetic variation should influence the potential for unisexuals to invade hermaphro-

dite populations lacking balanced polymorphisms. In hermaphrodite populations that are initially monomorphic at SA loci, selection may favor the invasion of haplotypes that carry unisexual sterility alleles and a rare complementary SA allele. Yet the waiting time until such double-mutant haplotypes arise may be long, particularly within small populations where mutational variation is limited (Weinreich and Chao 2005; Connallon and Clark 2010). However, while the origin and invasion of double-mutant haplotypes is improbable on generational timescales, such events become probable over the longer time intervals over which reproductive systems evolve. Consequently, such events may play a meaningful role in the evolution of dimorphic sexual systems. In addition, unisexual sterility alleles should readily invade hermaphrodite populations that are polymorphic for SA alleles. Such SA polymorphism could potentially segregate in hermaphrodite populations as balanced polymorphisms or as transient polymorphisms evolving under recurrent mutation, purifying selection, and genetic drift (see below).

Although we currently know very little about the extent of SA genetic variation in hermaphroditic species (e.g., Abbott 2011; Olito et al. 2018), several features of SA selection suggest that such variation may be common, particularly in large populations. First, theory suggests that both outcrossing and linkage between SA loci should promote the evolution of balanced SA polymorphisms in hermaphrodite populations, especially in heterogeneous environments (Jordan and Connallon 2014; Olito 2017; Olito et al. 2018). Second, SA alleles that are destined to eventually become fixed or lost are expected to segregate for many generations compared to unconditionally deleterious or beneficial alleles (Connallon and Clark 2012). The long transit times for SA alleles provide an opportunity for unisexual sterility mutations to arise on haplotypes bearing SA alleles that may ultimately be lost (e.g., Weinreich and Chao 2005). However, empirically identifying SA loci is challenging, even within dioecious species under strong SA selection, where few candidate loci have been identified (e.g., Barson et al. 2015; Lee et al. 2016; reviewed in Mank 2017; Connallon and Hall 2018). Further empirical attention to SA genetic variation in hermaphroditic species would help clarify the potential for linked SA selection to facilitate transitions to dioecy. An important caveat to our models of androdioecy and gynodioecy is that several mechanisms, including the evolution of sex-specific gene expression (Vicoso et al. 2013), may resolve the SA selection before the appearance of linked unisexual sterility mutations.

Mating Systems and the Evolution of Dioecy

The interplay between hermaphrodite mating systems and reproductive compensation is a key factor influencing the evolution of separate sexes in flowering plants, especially

via the gynodioecy pathway (Darwin 1877; Charlesworth and Charlesworth 1978a). Previous theory predicts that the evolution of gynodioecy is driven by the combination of reproductive compensation by unisexuals and inbreeding avoidance and is therefore most likely to occur in partially selfing populations (Lewis 1942; Lloyd 1975; Charlesworth and Charlesworth 1978a; Käfer et al. 2017). This prediction follows directly from the structure of equation (3), where \hat{k} depends entirely on the product of the selfing rate and inbreeding depression ($C\delta$). In contrast, the evolution of androdioecy is predicted to require significantly higher reproductive compensation, especially in partially selfing populations, because invading males must still compete with selfing hermaphrodites to fertilize ovules (Charlesworth and Charlesworth 1978b; Käfer et al. 2017).

The population selfing rate plays a critical role in our models as well, though our predictions differ from those of classical theory. The facilitating effect of linkage to an SA locus on the invasion of unisexual individuals is most prominent in outcrossing populations, and the effect of linkage weakens with increased selfing. Moreover, outcrossing hermaphrodite populations are more likely than selfing ones to maintain SA genetic variation (Jordan and Connallon 2014; Olito 2017). Both factors should promote evolutionary transitions from outcrossing hermaphrodite populations to androdioecious and gynodioecious systems. Thus, single-locus theory for the initial steps toward the evolution of dioecy may significantly underestimate the potential for evolutionary transitions to dimorphic sexual systems in predominantly outcrossing ancestral species. On the other hand, our models agree with previous theory that the conditions favoring transitions to androdioecy will remain difficult to satisfy when the ancestral rate of self-fertilization is high (large k ; eq. [6]; Charlesworth and Charlesworth 1978a). Hence, our results comport with prior theory in predicting that the evolution of androdioecy—and of dioecy via the androdioecy pathway—will remain more restrictive than transitions to gynodioecy (Charlesworth and Charlesworth 1978a; Charlesworth 2006; Renner 2014; Käfer et al. 2017).

Although the classic single-locus predictions for the evolution of androdioecy and gynodioecy are widely accepted, one of the key predictions—an association between the ancestral selfing rate and evolutionary transitions to dioecy—is not well supported empirically (Charlesworth 1985, 2006; Renner 2014). Our models can account for the weak association between the ancestral selfing rate and transitions to dioecy. In species where unisexual sterility alleles arise in linkage with an SA locus, evolutionary transitions to dimorphic sexual systems should be elevated in predominantly outcrossing hermaphrodite taxa. If transitions to dioecy involve a mixture of the traditional (i.e., single-locus) path and the alternative that we have proposed, then the net effect will be low (or no) correlation between the ancestral

hermaphrodite selfing rate and transitions to dioecy. In light of our results, it would be worth reexamining the association between the inferred pattern of ancestral self-fertilization and the transition rate between sexual systems among angiosperm species, using modern phylogenetic comparative methods. Such a study would further help to identify species where our proposed mechanism for the evolution of gynodioecy is most likely to have played a role (e.g., gynodioecious and dioecious species that appear to have evolved from outcrossing ancestors).

The Population Genetic Basis of Sex Chromosomes and Separate Sexes

Our results also have implications for the role of SA genetic variation during the early stages of sex chromosome evolution. The process of sex chromosome evolution into highly differentiated X and Y (or Z and W) chromosomes is thought to proceed by the following series of steps. First, a new sex-determination gene (or linked gene cluster) originates on an ordinary pair of autosomes. Second, SA alleles benefitting the heterogametic sex are predicted to accumulate in tight linkage with the sex-determining locus on the proto-Y or proto-W chromosome. Third, this linked SA variation generates selection for suppressed recombination between the proto-sex chromosomes. Fourth, the non-recombining sex chromosome degenerates as a consequence of recombination suppression. Finally, mechanisms of dosage compensation may evolve in response to the loss of functional genes on the degenerate sex chromosome (Rice 1987; Charlesworth 2002; Charlesworth et al. 2005; Bachtrog 2006; Qiu et al. 2013; Bachtrog et al. 2014).

In the classical theory for the evolution of dioecy (Charlesworth and Charlesworth 1978a), the initial step of sex chromosome evolution, as outlined above, coincides with the transition to full dioecy via the sequential invasion of complementary gender modifiers in a tightly linked region of the genome and complete repulsion linkage disequilibrium with one another (e.g., proto-Y and proto-X chromosomes carry female-sterility and male-sterility alleles, respectively; Charlesworth and Charlesworth 1978a). The tightly coupled pair of loci segregate, effectively, as a single sex-determining locus, which sets the stage for further sex chromosome differentiation by the steps outlined above.

Our theoretical results suggest an alternative—and previously unrecognized—sex chromosome evolutionary pathway. In contrast to the standard view, where sexual antagonism emerges after the origin of the sex-determination locus, our models suggest that SA polymorphism may represent the first step, rather than an intermediate step, in the evolution of heteromorphic sex chromosome systems. This role of SA genetic variation during the origin of sex chromo-

somes is reminiscent of recent theory for the turnover of established sex chromosomes (van Doorn and Kirkpatrick 2007, 2010). However, in our models, SA polymorphism precedes the origin of the sex determination system. Following the transition to dioecy, the proto-sex chromosomes are already primed for the evolution of recombination suppression and further differentiation (Charlesworth and Charlesworth 1978a; Rice 1987; Bachtrog 2006; Qiu et al. 2013). Given the crucial role of genetic linkage in nearly every step of sex chromosome evolution, it seems plausible that the amount of SA genetic variation present prior to the origin of distinct sexes may help to explain the extensive variation in sex chromosome differentiation that has been observed in angiosperms (Charlesworth 2002; Bachtrog et al. 2014; Renner 2014).

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Literature Cited

- Abbott, J. K. 2011. Intra-locus sexual conflict and sexually antagonistic genetic variation in hermaphroditic animals. *Proceedings of the Royal Society B* 278:161–169.
- Andersson, M. 1994. *Sexual selection*. Princeton University Press, Princeton, NJ.
- Ashman, T.-L., J. A. Tennesen, R. M. Dalton, R. Govindarajulu, M. H. Koski, and A. Liston. 2015. Multilocus sex determination revealed in two populations of gynodioecious wild strawberry, *Fragaria vesca* subs. *bracteata*. *G3: Genes, Genomes, Genetics* 5:2759–2773.
- Bachtrog, D. 2006. A dynamic view of sex chromosome evolution. *Current Opinion in Genetics and Development* 16:578–585.
- Bachtrog, D., J. E. Mank, C. L. Peichel, M. Kirkpatrick, S. P. Otto, T.-L. Ashman, M. W. Hahn, et al. 2014. Sex determination: why so many ways of doing it? *PLoS Biology* 12:e1001899.
- Barrett, S. C. H. 2010. Darwin's legacy: the forms, function, and sexual diversity of flowers. *Philosophical Transactions of the Royal Society B* 365:351–368.
- Barson, N. J., T. Ayanat, K. Hindar, M. Baranski, G. H. Bolstad, P. Fiske, C. Jacq, et al. 2015. Sex-dependent dominance at a single locus maintains variation in age at maturity in salmon. *Nature* 528:405–408.
- Bawa, K. F. 1980. Evolution of dioecy in flowering plants. *Annual Review of Ecology and Systematics* 11:15–39.
- Blackburn, G. S., A. Y. K. Albert, and S. P. Otto. 2010. The evolution of sex ratio adjustment in the presence of sexually antagonistic selection. *American Naturalist* 176:264–275.
- Bull, J. J. 1983. *Evolution of sex determining systems*. Benjamin/Cummings, Menlo Park, CA.
- Charlesworth, B., and D. Charlesworth. 1978a. A model for the evolution of dioecy and gynodioecy. *American Naturalist* 112:975–997.
- . 2010. *Elements of evolutionary genetics*. Roberts, Greenwood Village, CO.
- Charlesworth, B., T. Miyo, and H. Borthwick. 2007. Selection responses of means and inbreeding depression for female fecundity in *Drosophila melanogaster* suggest contributions from intermediate-frequency alleles to quantitative trait variation. *Genetics Research* 89: 85–91.
- Charlesworth, D. 1985. Distribution of dioecy and self-incompatibility in angiosperms. Pages 237–268 in P. J. Greenwood and M. Slatkin, eds. *Evolution: essays in honor of John Maynard Smith*. Cambridge University Press, Cambridge.
- . 1999. Theories of the evolution of dioecy. Pages 33–60 in M. A. Geber, T. E. Dawson, and L. F. Delph, eds. *Gender and sexual dimorphism in flowering plants*. Springer, Heidelberg.
- . 2002. Plant sex determination and sex chromosomes. *Heredity* 88:94–101.
- . 2006. Evolution of plant breeding systems. *Current Biology* 16:R726–R735.
- Charlesworth, D., and B. Charlesworth. 1978b. Population genetics of partial male-sterility and the evolution of monoecy and dioecy. *Heredity* 41:137–153.
- Charlesworth, D., B. Charlesworth, and G. Marais. 2005. Steps in the evolution of heteromorphic sex chromosomes. *Heredity* 95:118–128.
- Charlesworth, D., and J. Willis. 2009. The genetics of inbreeding depression. *Nature Reviews Genetics* 10:783–796.
- Connallon, T., and A. G. Clark. 2010. Sex linkage, sex-specific selection, and the role of recombination in the evolution of sexually dimorphic gene expression. *Evolution* 64:3417–3442.
- . 2012. A general population genetic framework for antagonistic selection that accounts for demography and recurrent mutation. *Genetics* 190:1477–1489.
- Connallon, T., and M. D. Hall. 2018. Genetic constraints on adaptation: a theoretical primer for the genomics era. *Annals of the New York Academy of Sciences* 1422:65–87.
- Connallon, T., and C. Y. Jordan. 2016. Accumulation of deleterious mutations near sexually antagonistic genes. *G3: Genes, Genomes, Genetics* 6:2273–2280.
- Crnokrak, P., and S. C. H. Barrett. 2002. Purging the genetic load: a review of the experimental evidence. *Evolution* 56:2347–2358.
- Darwin, C. 1877. *The different forms of flowers on plants of the same species*. J. Murray, London.
- Dufay, M., and E. Billard. 2012. How much better are females? the occurrence of female advantage, its proximal causes and its variation within and among gynodioecious species. *Annals of Botany* 109:505–519.
- Frank, S. A. 1989. The evolutionary dynamics of cytoplasmic male sterility. *American Naturalist* 133:345–376.
- Fry, J. D. 2009. The genomic location of sexually antagonistic variation: some cautionary comments. *Evolution* 64:1510–1516.
- Garcia-Dorado, A. 2017. An explicit model for the inbreeding load in the evolutionary analysis of selfing. *Evolution* 71:1381–1389.

- Goldberg, E. E., S. P. Otto, J. C. Vamosi, I. Mayrose, N. Sabath, R. Ming, and T.-L. Ashman. 2017. Macroevolutionary synthesis of flowering plant sexual systems. *Evolution* 71:898–912.
- Gregorious, H.-R. 1982. Selection in plant populations of effectively infinite size II. Protectedness of a biallelic polymorphism. *Journal of Theoretical Biology* 96:689–705.
- Hedrick, P. W., and A. Garcia-Dorado. 2016. An explicit model for the inbreeding load in the evolutionary analysis of selfing. *Trends in Ecology and Evolution* 31:940–952.
- Husband, B. C., and D. W. Schemske. 1996. Evolution of the magnitude and timing of inbreeding depression in plants. *Evolution* 50:54–70.
- Jarne, P., and J. R. Auld. 2006. Animals mix it up too: the distribution of self-fertilization among hermaphroditic animals. *Evolution* 60:1816–1824.
- Jordan, C. Y., and T. Connallon. 2014. Sexually antagonistic polymorphism in simultaneous hermaphrodites. *Evolution* 68:3555–3569.
- Käfer, J., G. A. B. Marais, and J. R. Pannell. 2017. On the rarity of dioecy in flowering plants. *Molecular Ecology* 26:1225–1241.
- Kidwell, J. F., M. T. Clegg, F. M. Stewart, and T. Prout. 1977. Regions of stable equilibria for models of selection in the two sexes under random mating. *Genetics* 85:171–183.
- Lande, R., and E. Porcher. 2017. Inbreeding depression maintained by recessive lethal mutations interacting with stabilizing selection on quantitative characters in a partially self-fertilizing population. *Evolution* 71:1191–1204.
- Lande, R., and D. W. Schemske. 1985. The evolution of self-fertilization and inbreeding depression in plants. I. Genetic models. *Evolution* 39:24–40.
- Lee, Y. W., L. Fishman, J. K. Kelly, and J. H. Willis. 2016. A segregating inversion generates fitness variation in yellow monkeyflower (*Mimulus guttatus*). *Genetics* 202:1473–1484.
- Lewis, D. 1941. Male sterility in natural populations of hermaphrodite plants. *New Phytologist* 40:56–63.
- . 1942. The evolution of sex in flowering plants. *Biological Reviews* 17:46–67.
- Lloyd, D. G. 1975. The maintenance of gynodioecy and androdioecy in angiosperms. *Genetica* 45:325–339.
- . 1976. The transmission of genes via pollen and ovules in gynodioecious angiosperms. *Theoretical Population Biology* 9:299–316.
- Mank, J. E. 2017. Population genetics of sexual conflict in the genomic era. *Nature Reviews Genetics* 18:721–730.
- Morgan, M. T. 1992a. The evolution of traits influencing male and female fertility in outcrossing plants. *American Naturalist* 139:1022–1051.
- . 1992b. Attractive structures and the stability of hermaphroditic sex expression in flowering plants. *Evolution* 46:1199–1213.
- Ohta, T., and C. C. Cockerham. 1974. Detrimental genes with partial selfing and effects on a neutral locus. *Genetics Research* 23:191–200.
- Olito, C. 2017. Consequences of genetic linkage for the maintenance of sexually antagonistic polymorphism in hermaphrodites. *Evolution* 71:458–464.
- Olito, C., J. K. Abbott, and C. Y. Jordan. 2018. The interaction between sex-specific selection and local adaptation in species without separate sexes. *Philosophical Transactions of the Royal Society B* 373:20170426.
- Otto, S., and T. Day. 2007. *A biologist's guide to mathematical modeling in ecology and evolution*. Princeton University Press, Princeton, NJ.
- Patten, M., D. Haig, and F. Úbeda. 2010. Fitness variation due to sexual antagonism and linkage disequilibrium. *Evolution* 64:3638–3642.
- Prout, T. 2000. How well does opposing selection maintain variation? Pages 157–203 in R. S. Singh and C. B. Kimbras, eds. *Evolutionary genetics from molecules to morphology*. Cambridge University Press, Cambridge.
- Qiu, S., R. Bergero, and D. Charlesworth. 2013. Testing for the footprint of sexually antagonistic polymorphisms in the pseudoautosomal region of a plant sex chromosome pair. *Genetics* 194:663–672.
- Renner, S. 2014. The relative and absolute frequencies of angiosperm sexual systems: dioecy, monoecy, gynodioecy, and an updated online database. *American Journal of Botany* 101:1588–1596.
- Rice, W. R. 1987. The accumulation of sexually antagonistic genes as a selective agent promoting the evolution of reduced recombination between primitive sex chromosomes. *Evolution* 41:911–914.
- Roze, D. 2015. Effects of interference between selected loci on the mutation load, inbreeding depression, and heterosis. *Genetics* 201:745–757.
- Sakai, A. K., and S. G. Weller. 1999. Gender and sexual dimorphism in flowering plants. Pages 1–31 in M. A. Geber, T. E. Dawson, and L. F. Delph, eds. *Gender and sexual dimorphism in flowering plants*. Springer, Heidelberg.
- Seger, J., and V. M. Eckhart. 1996. Evolution of sexual systems and sex allocation in plants when growth and reproduction overlap. *Proceedings of the Royal Society B* 263:833–841.
- Úbeda, F., D. Haig, and M. Patten. 2010. Stable linkage disequilibrium owing to sexual antagonism. *Proceedings of the Royal Society B* 278:855–862.
- van Doorn, G. S., and M. Kirkpatrick. 2007. Turnover of sex chromosomes induced by sexual conflict. *Nature* 449:909–912.
- . 2010. Transitions between male and female heterogamety caused by sex-antagonistic selection. *Genetics* 186:629–645.
- Vicoso, B., V. B. Kaiser, and D. Bachtrog. 2013. Sex-biased gene expression at homomorphic sex chromosomes in emus and its implication for sex chromosome evolution. *Proceedings of the National Academy of Sciences of the USA* 110:6453–6458.
- Webb, C. J. 1999. Empirical studies: evolution and maintenance of dimorphic breeding systems. Pages 61–95 in M. A. Geber, T. E. Dawson, and L. F. Delph, eds. *Gender and sexual dimorphism in flowering plants*. Springer, Heidelberg.
- Weinreich, D. M., and L. Chao. 2005. Rapid evolutionary escape by large populations from local fitness peaks is likely in nature. *Evolution* 59:1175–1182.
- Westergaard, M. 1958. The mechanism of sex determination in dioecious flowering plants. *Advances in Genetics* 9:217–281.

References Cited Only in the Online Appendixes

- Caballero, A., and W. G. Hill. 1992. Effects of partial inbreeding on fixation rates and variation of mutant genes. *Genetics* 131:493–507.
- Connallon, T., and A. Clark. 2014. Balancing selection in species with separate sexes: insights from Fisher's geometric model. *Genetics* 197:991–1006.
- Nagylaki, T. 1997. The diffusion model for migration and selection in a plant population. *Journal of Mathematical Biology* 35:409–431.

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Correction

Shortly after publication of “Sexually Antagonistic Variation and the Evolution of Dimorphic Sexual Systems” by Colin Olito and Tim Connallon (*American Naturalist* 193:688–701), the authors noticed a mistake in the analytic results pertaining to the invasion of sexually antagonistic alleles (hereafter, “SA alleles”) linked to a unisexual sterility allele.

In the section titled “Invasion of Unisexuals into Monomorphic Populations,” we described how linkage to a dominant unisexual sterility allele expands invasion conditions of an SA allele relative to single-locus expectations (eqq. [5] and [7] and surrounding paragraphs). These results mistakenly conditioned on the single-locus invasion threshold for the sterility allele being met exactly. Specifically, we substituted $k = 1 - 2C\delta$ from equation (3) when deriving equation (5) as well as $k = (1 + C(1 - 2\delta))/(1 - C)$ from equation (6) when deriving equation (7). Equations (5) and (7) are mathematically correct for these conditions, but they do not provide general expressions for when the two-locus invasion conditions for SA alleles become more permissive than single-locus predictions. Below we briefly present the correct general condition for each model under the assumptions described in the article. To avoid confusion, we reference equations from the article by their original numbers and the corrected equations with lowercase roman numerals.

Generalization of equation (5). Under arbitrary levels of selfing and inbreeding depression ($0 \leq C, \delta \leq 1$), the conditions for invasion of a haplotype linking a codominant ($h_f = h_m = 1/2$) female-benefit SA allele and dominant male-sterility allele (AM_2 in the gynodioecy model, corresponding to $\lambda_{AM} > 1$) are more permissive than the single-locus SA invasion condition for female-beneficial allele invasion when

$$r < \frac{2 - s_m - k(2 + s_m) + C(2 + s_m + k(s_m + 4\delta - 2) - 8\delta) - 4C^2\delta(1 - 2\delta)}{C(1 + k)(s_m + 4\delta - 2) - (1 + k)(2 + s_m)}. \quad (\text{i})$$

Equation (i) shows that greater reproductive compensation, inbreeding depression, and selection against the male-benefit allele (higher k , δ , and s_m) all act to reduce the threshold level of linkage that expands invasion conditions for the female-benefit allele (the critical r in eq. [i] increases), whereas increased selfing (higher C) increases the requisite level of linkage (the critical r decreases). We can confirm our mistake by substituting $k = 1 - 2C\delta$ (from eq. [3]) into equation (i) and simplifying, which recovers our original equation (5).

Generalization of equation (7). For the androdioecy model, invasion of a haplotype linking the male-benefit SA allele and a female-sterility allele (aM_2 , corresponding to $\lambda_{AM} > 1$) is more permissive than the single-locus SA invasion criterion when

$$r < \frac{2 - s_f - k(2 + s_f) + C(1 + k)(2 - s_f) + 2C\delta(s_f(1 + k) - 2)}{(1 + k)(C(2 - s_f(1 - 2\delta)) - 2 - s_f)}. \quad (\text{ii})$$

In equation (ii), greater reproductive compensation, inbreeding depression, and selection against the female-benefit allele (higher k , δ , and s_f) all reduce the threshold level of linkage that expands the two-locus invasion conditions for the male-benefit allele (the critical r in eq. [ii] increases), whereas increased selfing increases the requisite level of linkage (the critical r decreases). Substituting $k = (1 + C(1 - 2\delta))/(1 - C)$ (from eq. [6]) into equation (ii) and simplifying recovers our original equation (7), confirming the mistake. This error may cause confusion because we emphasized in the article that increased selfing expands the conditions for invasion of a male-beneficial SA allele in the androdioecy model. While this is true under the limited conditions imposed by the substitution, the critical level of recombination described by equation (ii) declines with increased selfing (higher C).

We made the same error in appendix C when deriving results for a dominance reversal at the SA locus ($h_f = h_m = 1/4$; eqq. [C2], [C3]). The correct expression for the gynodioecy model under dominance reversal and obligate outcrossing ($C = 0$) is

$$r < 1 - \frac{8}{(1 + k)(4 + s_m)}, \quad (\text{iii})$$

which should replace equation (C2). The corresponding result for the model of androdioecy is identical to equation (iii), except with s_f replacing s_m , and should replace equation (C3).

We note that these errors affect neither the main results of the article nor its conclusions. We apologize for any confusion our mistakes may have caused.

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